



Full Length Research Paper

## PARASITIC FAUNA OF AMPHIBIANS FROM RUMUJI-EMOHUA, RIVERS STATE, NIGERIA.

Amuzie\*<sup>1</sup>, Chidinma Charity, Agala<sup>1</sup>, King Abraham and Aisien<sup>2</sup>, Martins Siaka

<sup>1</sup>Department of Animal and Environmental Biology, Rivers State University of Science and Technology, P.M.B. 5080, Nkpolu-Oroworukwo, Rivers State, Nigeria.

<sup>2</sup>Laboratory of Parasitology Research, Department of Animal and Environmental Biology, University of Benin, P.M.B. 1154, Benin City, Nigeria.

\*Corresponding Author's E-Mail: [ekeh.chidinma@ust.edu.ng](mailto:ekeh.chidinma@ust.edu.ng)

### ABSTRACT

A survey of the parasites of amphibians from Rumuji in Emohua Local Government Area of Rivers State, Nigeria, was conducted in the months of April and May, 2016. Anurans collected included *Amietophrynus maculatus*, *A. cameroonensis*, *Hoplobatrachus occipitalis*, *Ptychadena oxyrynchus* and *Pty. pumilio*. Parasites recovered included a *Raillietiella* sp. (Pentastomida): *Cylindrotaenia jaegerskioeldi* (Cestoda), *Mesocoelium monodi* and *Diplodiscus fischthalicus* (Digenea); *Rhabdias africanus*, *Rhabdias* sp., *Amplicaeum africanum*, *Oswaldocruzia hoepplii*, *Cosmocerca ornata*, and a larval ascaridoid (Nematoda). The co-occurrence of *Raillietiella* sp. and *R. africanus* in the lungs of *A. maculatus* is being reported for the first time in Nigeria. *Amietophrynus maculatus* is a new host record for *D. fischthalicus* in the country. The occurrence of *Oswaldocruzia hoepplii* in *A. cameroonensis* also increases the host range of the parasite in Nigerian anurans.

**Key words:** Amphibians, Parasites, Emohua, Rivers State, Nigeria.

### INTRODUCTION

Amphibians are ectothermic, tetrapod vertebrates of the class Amphibia which exhibit a biphasic mode of life. They inhabit a wide variety of habitats, with most species living within terrestrial, fossorial, arboreal and freshwater aquatic ecosystems (Blackburn and Wake, 2011). They serve as paratenic or definitive hosts to a number of parasites (Imasuen *et al.*, 2012), most of which are acquired from the consumption of intermediate hosts such as beetles and snails.

The Niger Delta is home to several amphibian species on which very limited

studies have been undertaken (Akani *et al.*, 2004; Ogoannah, 2010; Ogoannah *et al.*, 2014). Urgent attention needs to be focused on this task in view of the numerous oil prospecting activities in this region and the attendant environmental pollution and degradation associated with them. These activities negatively impact on amphibian diversity and health (Akani *et al.*, 2004). Unlike other bioclimatic zones of Nigeria where much information is available on the parasitic infections of amphibians hosts (Oladimeji *et al.*, 1990; Aisien *et al.*, 2001, 2003, 2004a, b, 2009; 2015a,b, Imasuen and Aisien, 2012; Imasuen *et al.*, 2012; Nworah and Olorunfemi, 2013, Edo-Taiwo *et al.*, 2014)

studies on this subject from the Niger Delta is still limited (Anyacho, 1997; Aisien *et al.*, 2001; Oddo, 2008; Akani *et al.*, 2011; Omereji, 2014; Sampson, 2014).

In view of the paucity of information on the amphibians of the Niger Delta and their endo-parasite fauna, there is a need to urgently fill this lacuna by undertaking relevant studies which will add to the pool of emerging information on this subject. The present study was therefore conducted to examine the amphibian diversity and parasitic infections of anurans from Rumuji in Emohua Local Government Area of Rivers State of Nigeria, results of which we present in this paper.

## MATERIALS AND METHODS

The study area is located in Rumuji community (N4°56'32'', E6° 46'57'') in Emohua Local Government Area of Rivers State, Nigeria. It is located within the rainforest zone, with characteristic tropical climatic conditions. The vegetation in the community is composed of sparse secondary vegetation which included standing bushes and grass fields along with some nearby cassava farms. These were searched for the presence and collection of amphibians. Searches were also conducted along tracks on the road. The survey was carried out during the period spanning the last week of April, 2016 to the end of May, 2016. This was the early rainy season characterized by very few rainy days with much of sunshine and heat.

Adult frogs were hand captured, at night, between 7 pm and 11 pm, using the Visual Encounter and Acoustic Survey method (Crump and Scott, 1994). Individual frogs were transported live to the laboratory in moistened plastic bottles with perforated lids. Host specimens were identified (Roedel, 2000) and killed by exposure to chloroform vapour in an airtight jar. The sexes of the anurans were determined by examining for the presence of vocal sacs, and examining the sex

organs after they were dissected. The frogs were dissected and the oesophagus/stomach, small intestine, large intestine/rectum, lungs, urinary bladder were isolated and transferred to Petri dishes containing 0.72% NaCl solution and examined for parasites with a dissecting microscope. The body cavity was also examined for encysted parasites. Nematodes were fixed in hot 70% alcohol and preserved in fresh 70% alcohol. Cestodes and trematodes were flattened under cover slip pressure on a microscope slide and fixed in 5% formol-saline. The fixed specimens were preserved in the same fixative in labelled specimen bottles. Nematode parasites were cleared in lactophenol before examination. Cestodes and trematodes were washed free of fixative in three to four changes of tap water at 45mins intervals and then stained overnight in a dilute solution of acetocarmine. The specimens were dehydrated in alcohol series and permanent mounts made in Canada balsam. Parasites were identified with the protocols of Yamaguti (1971), Prudhoe and Bray (1982) and Khalil *et al.* (1994).

## STATISTICAL ANALYSIS

The critical parasite quantification measures, namely prevalence and mean intensity, were computed according to Bush *et al.* (1997). The Data were  $\log(x+1)$  transformed (Zar, 1984) and subjected to t-tests to determine significant differences in prevalence rates of the parasite species in infected hosts between locations. Standard deviation was computed for the mean intensity values. Statistical analyses were performed using Microsoft Excel. Kruskal-Wallis test was used to test for significant differences between parasite burdens in the predilection sites (Fowler and Cohen, 1994).

## RESULTS

A total of sixty-one (61) anurans (15 females and 46 males) consisting predominantly of *Amietophrynus*

*maculatus* (38) and *Hoplobatrachus occipitalis* (15) were examined for parasites. Others included *A. cameroonensis* (1), *Ptychadena*

*oxyrhynchus* (2) and *Ptychadena pumilio* (5). The host species abundance during the sampling period is shown in Table 1.

**Table 1. Relative abundance of amphibian hosts in Rumuji, Rivers State**

Host	Number examined	Abundance (%)	Males	Females
<i>A. cameroonensis</i>	1	1.64	0	01
<i>A. maculatus</i>	38	42.62	27	11
<i>H. occipitalis</i>	15	3.28	14	01
<i>P. oxyrhynchus</i>	2	1.64	02	00
<i>P. pumilio</i>	5	3.28	03	02
<b>Total</b>	<b>61</b>	<b>100</b>	<b>46</b>	<b>15</b>

The sites of infection in the anurans investigated are presented in Table 2; the small intestine was the most parasitized site. The parasites recovered from the anurans included a pentastomid (*Raillietiella* sp.), one cestode (*Cylindrotaenia jaegerskioeldi*), two digenetic trematodes (*Mesocoelium monodi* and *Diplodiscus fischthalicus*). Nematodes were also recovered and they included *Rhabdias africanus*, an undetermined *Rhabdias* sp., *Amplificaecum africanum*, *Oswaldocruzia hoepplii*, *Cosmocerca ornata* and a larval ascaridoid encysted in the body cavity. A co-occurrence of the *Raillietiella* sp. and *R. africanus* was recorded in the lungs of two specimens of *A. maculatus* in a ratio of 2:9 in the first specimen and 26:2 in the other.

The prevalence of the parasites found ranged from 2.63% to 100.00%, while the mean intensity of infection ranged from 1.00 to 11.57 parasites/infected host (Table 3). The overall prevalence of infection in male and female amphibians is presented in Table 4. The overall prevalence of parasites in female frogs was higher (80%) than that of the males (57.8%) as shown in Tables 4, but the difference was statistically insignificant ( $t_{26} = 0.256$ ,  $p = 0.800$ ). *Amietophrynus cameroonensis* was

represented by female hosts while *Pty. oxyrhynchus* and *H. occipitalis* were represented by male hosts. In those anuran species represented by both sexes (*A. maculatus*, *P. pumilio*), parasite prevalence was higher in female hosts (91:81 for *A. maculatus*; 50:33.3 for *P. pumilio*). In *A. maculatus* which was the dominant species in the amphibian collection from Rumuji, *D. fischthalicus* and the larval ascaridoid infections were recorded only in male toads while the other parasites were encountered in both sexes (Table 5). While the prevalence values were higher in males for *R. africanus*, *A. africanum* and *C. ornata*, higher values were recorded for the *Raillietiella* sp., *M. monodi* and *O. hoepplii* in female hosts. Of the 15 *H. occipitalis* examined, *C. ornata* occurred only on male frogs (Table 5). 99% confidence levels with a Kruskal-Wallis test showed a highly significant difference ( $K = 71.32$ ,  $p = 0.1$ ) between the predilection sites. The non-parametric Tukey-type multiple comparisons test revealed that significant differences existed between the helminth burden in the intestine and oesophagus/stomach, and between the large intestine/colon and the oesophagus stomach. However, significant differences did not exist between the other comparisons made.

**Table 2. Endoparasites of anurans from Rumuji, Rivers State, Nigeria**

<b>Parasites</b>	<b>Hosts</b>	<b>Site of infection</b>
<b>Pentastomida</b>		
<i>Railietiella</i> sp.	<i>A. cameroonensis</i>	Lungs
	<i>A. maculatus</i>	Lungs
<b>Cestoda</b>		
<i>C. jaegerskioeldi</i>	<i>P. pumilio</i>	Small intestine
<b>Trematoda</b>		
<i>D. fischthalicus</i>	<i>A. maculatus</i>	Rectum
<i>M. monodi</i>	<i>A. maculatus</i>	Small intestine
	<i>P. oxyrynchus</i>	Small intestine
<b>Nematoda</b>		
<i>A. africanum</i>	<i>A. maculatus</i>	Small intestine
	<i>P. pumilio</i>	Small intestine
<i>R. africanus</i>	<i>A. maculatus</i>	Lungs
<i>Rhabdias</i> sp	<i>P. pumilio</i>	Lungs
<i>O. hoepplii</i> .	<i>A. maculatus</i>	Small intestine
<i>C. ornata</i>	<i>A. maculatus</i>	Rectum
	<i>H. occipitalis</i>	Rectum
Larval ascaridoid	<i>A. cameroonensis</i>	Body cavity
	<i>A. maculatus</i>	Body cavity

**Table 3: Prevalence of parasitic infections in the anuran host species from Rumuji, Rivers State**

<b>Parasite</b>	<b>Host</b>	<b>Prevalence (%)</b>	<b>*Mean intensity</b>
<b>Pentastomida</b>			
<i>Railietiella</i> sp.	<i>A. cameroonensis</i>	100.0	15.0
	<i>A. maculatus</i>	18.42	11.6±4.43
<b>Cestoda</b>			
<i>C. jaegerskioeldi</i>	<i>P. pumilio</i>	20.0	1.0±0.00
<b>Trematoda</b>			
<i>D. fischthalicus</i>	<i>A. maculatus</i>	2.6	11.0±0.00
<i>M. monodi</i>	<i>P. oxyrhynchus</i>	50.0	9.0±3.25
	<i>A. maculatus</i>	18.4	6.6±5.41
<b>Nematoda</b>			
<i>A. africanum</i>	<i>A. maculatus</i>	29.0	1.8±1.61
	<i>P. pumilio</i>	20.0	2.0±0.00
<i>R. africanus</i>	<i>A. maculatus</i>	15.8	3.2±3.25
<i>Rhabdias</i> sp.	<i>P. pumilio</i>	20.0	1.0±0.00
<i>O. hoepplii</i>	<i>A. maculatus</i>	42.1	4.6±4.55
Larval ascaridoid	<i>A. cameroonensis</i>	100	2.0±0.00
	<i>A. maculatus</i>	5.3	1.5±1.41
<i>C. ornata</i>	<i>A. maculatus</i>	34.2	6.1±5.45
	<i>H. occipitalis</i>	13.3	2.0±0.00

**Table 4: Overall prevalence of helminth parasites according to sex in the amphibians examined**

Hosts	Male amphibians		Female amphibians	
	No. examined	No. infected	No. examined	No. Infected
<i>A. maculatus</i>	27	22	11	10
<i>A. cameroonensis</i>	0	0	1	1
<i>H. occipitalis</i>	14	2	0	0
<i>P. oxyrhynchus</i>	2	1	0	0
<i>P. pumilio</i>	3	1	2	1
Total	45	26	15	12

**Table 5. Parasite prevalence and mean intensity in the amphibian hosts from Rumuji, Rivers State according to sex**

Host	Parasite	Males		Females	
		P	MI	P	MI
<i>A. cameroonensis</i>	<i>Railietiella</i> sp.	-	-	100	15.0
<i>A. maculatus</i>	<i>Railietiella</i> sp.	14.8	1.8	27.3	24.7
	<i>D. fischthalicus</i>	3.7	11.0	-	-
	<i>M. monodi</i>	11.1	5.0	36.4	7.8
	<i>R. africanus</i>	22.2	3.0	9.1	2.0
	<i>A. africanum</i>	37.0	2.6	9.1	1.0
	<i>O. hoeplii</i>	37.0	3.3	54.6	6.8
	<i>C. ornata</i>	40.7	6.1	18.2	9.5
	Larval ascaridoid	7.4	1.5	-	-
	<i>P. oxyrhynchus</i>	<i>M. monodi</i>	50.0	9.0	-
<i>P. pumilio</i>	<i>C. jaegerskioeldi</i>	33.3	1.0	-	-
	<i>A. africanum</i>	-	-	50.0	2.0
	<i>Rhabdias</i> sp.	-	-	50.0	1.0
<i>H. occipitalis</i>	<i>C. ornata</i>	14.3	2.0	-	-

## DISCUSSION

The amphibian species encountered at Rumuji were few when compared to the 11 recorded at Egbeda (Emuohua) by Omereji (2014). Whereas Omereji (2014) recorded five tree frog species at Egbeda, these frogs were not encountered during the sampling period at Rumuji. This may be attributable to the short duration of the study, the nature of the sampling sites which consisted mainly of secondary forest and the season of collection which was the early part of the rainy season. Nevertheless, the number of parasite species (10) recovered from the anurans from Rumuji is quite comparable to the 12 reported from the Egbeda anurans.

Pentastomids (*Railietiella* spp.) are known lung parasites of

*Amietophrynus regularis* (Aisien *et al.*, 2001; Ozemoka, 2012; Sampson, 2014) and a *Hyperolius* sp. (Ovwah, 2016). In this study, these parasites were recovered from *A. cameroonensis* and *A. maculatus*, which are new host records for these parasites. In contrast to the observation from previous studies, where these parasites were reported to occur separately in the lungs of infected hosts, we observed their co-occurrence with *R. africanus* (nematode) in the lungs of *A. maculatus*. This is the first report of such an occurrence in anurans of Nigeria.

The only cestode recovered in this study was *Cylindrotaenia jaegerskioeldi* from the small intestine of *Ptychadena pumilio*. This parasite is a multi-host parasite having been reported in *A.*

*regularis* and *A. maculatus* (Aisien *et al.*, 2001, 2003, 2004), *Ptychadena* spp. (Aisien *et al.*, 2003, 2009), *Aubria subsigilata* (Aisien *et al.*, 2009) and tree frogs (Imasuen *et al.*, 2012; Sampson, 2014). Sampson (2014) reported 52.2% prevalence of this parasite in the tree frog, *Afrixalus fulvovittatus*, which was attributed to the altered nature of the study environment in urban Port Harcourt. *Cephalochlamys compactus*, another cestode infecting *H. occipitalis* in locations in the Niger Delta (Aisien *et al.*, 2001; Omereji, 2014) was not recorded in the *H. occipitalis* investigated at Rumuji.

It is surprising that only two trematodes (*D. fischthalicus* and *M. monodi*) were recorded in the anurans from Rumuji. In an earlier study by Omereji (2014) in two locations in Rivers State (Egbeda in Emohua and Odioku in Ahoada), five trematodes (*Mesocoelium monodi*, *Haematoloechus exoterorchis*, *Ganeo africana*, *Prosotochus exovitellosus* and *Progonimodiscus doyeri*) were recorded. Three of these parasites (*Haematoloechus exoterorchis*, *Ganeo africana*, *Prosotochus exovitellosus*) occurred in *H. occipitalis*, one (*P. doyeri*) in *Hymenochirus* sp. while *M. monodi* infected *Ptychadena* spp. (*P. bibroni* and *P. oxyrhynchus*). *Mesocoelium monodi* and *Diplodiscus fischthalicus* also infect anuran hosts elsewhere in Nigeria. *Mesocoelium monodi* has been recorded in several amphibian hosts including *Amietophrynus regularis*, *A. maculatus*, *Chiromantis rufescens*, *Hoplobatrachus occipitalis*, *Phrynobatrachus* sp., and in *Ptychadena* spp. (Aisien *et al.*, 2001, 2003, 2004, 2009; Imasuen and Aisien, 2012; Imasuen *et al.*, 2012). Whereas *D. fischthalicus* is already known to infect *H. occipitalis*, *R. galamensis*, *A. subsigillata* and *P. pumilio* (Aisien *et al.*, 2001, 2003, 2004, 2009; Imasuen and Aisien, 2012; Imasuen *et al.*, 2012), in *Phrynobatrachus francisci* (Omereji, 2014), this is the first record of *D. fischthalicus* in *A. maculatus*,

thus extending the host range of the parasite.

All the nematodes recorded in this study have previous records in anurans investigated elsewhere in Nigeria (Aisien *et al.*, 2001, 2003, 2004; Imasuen *et al.*, 2012). *Rhabdias africanus* infects *Amietophrynus* spp. (Kuzmin, 2001) but there are reported occurrences of this lung worm in other anuran hosts that needs to be confirmed. This has become imperative with the recovery of *Rhabdias* spp. with features different from those of *R. africanus*. The *Rhabdias* sp. recovered from the lungs of a *Pty. pumilio* in this study and those from other anurans belong to this group. *Amplificaecum africanum* has been recovered from *A. regularis*, *A. maculatus*, and *H. occipitalis* (Aisien *et al.*, 2001, 2003) while *O. hoeplli* commonly infects *Amietophrynus* and *Ptychadena* spp. *Amietophrynus cameroonensis* thus becomes a new host record for this nematode. *Cosmocerca ornata* has been reported from several hosts including *Amietophrynus* sp., *H. occipitalis*, *Ptychadena* spp., *Xenopus muelleri*, *Afrixalus dorsalis*, *Hyperolius fusciventris burtoni* and *H. hyloides* (Aisien *et al.*, 2001; Imasuen *et al.*, 2012). Encysted larval ascaridoids found in *Amietophrynus* spp. in this study have previously been reported in the body cavity of several anuran species (Aisien *et al.*, 2001, 2003, 2004, 2009, 2015) and according to Imasuen *et al.* (2012) these nematodes use the amphibians as paratenic hosts to reach their definitive hosts.

It is not possible to comment on the parasite prevalence in those anuran hosts represented by only one of the sexes. However, in those species represented by both sexes (*A. maculatus*, *P. pumilio* and *H. occipitalis*), infection prevalence was consistently higher in the female hosts. Although this observation was not statistically significant, it may be related to their feeding habits. Gravid female frogs tend to forage more for food in order to satisfy their nutritional and those of the

developing egg mass. In this process, they consume more infected invertebrate intermediate hosts than female frogs.

## CONCLUSION

In conclusion, this study has examined the parasitic infections of anurans from Rumuji-Emohua in Rivers State of Nigeria. Although all the parasites recorded in this work have been reported in anurans elsewhere in Nigeria, there are certain peculiarities that are noteworthy in this location. We reported for the first time in Nigerian anurans, the co-occurrence of pentastomids and *R. africanus* in the lungs of *A. maculatus*. This toad is also reported here as a new host record for *D. fischthalicus*. Pentastomids and *O. hoepflii* are also being reported new parasite records in *A. cameroonensis*. This study has added new information to the existing database on the amphibians of Rivers State and their parasitic infections.

## REFERENCES

- Aisien, S.O., Ugbo, A.D., Ilavbare, A.N. and Ogunbor, O. (2001). Endoparasites of amphibians from south-western Nigeria. *Acta Parasitologica*, 46: 299-305.
- Aisien, S.O., Ajakaiye, F.B. and Braimoh, K. (2003). Helminth parasites of anurans from the savannah-mosaic zone of south-western Nigeria. *Acta Parasitologica*, 48: 47-54.
- Aisien, S.O., Ayeni, F. and Ilechie, I. (2004a). Helminth fauna of anurans from the Guinea savanna at New Bussa, Nigeria. *African Zoology*, 39: 133-136.
- Aisien, S.O., Salami, L.A., Obaro, F.E. and Erakpoweri, S.O. (2004b). The influence of climate on the distribution of monogeneans of anurans in Nigeria. *Journal of Helminthology*, 78: 101-104.
- Aisien, S.O., Ogoannah, S.O., and Imasuen, A.A. (2009). Helminth parasites of amphibians from a rainforest reserve in south-western Nigeria. *African Zoology* 44: 1-7.
- Aisien, M.S.O., Aigbirior, P.O., Ovwah, E., Edo-Taiwo, O. (2015a). Blood parasites of some anurans from Edo State, Nigeria. *Tropical Biomedicine* 32(4): 598-607.
- Aisien, M.S.O., Uwagbae, M., Edo-Taiwo, O., Imasuen, A.A. and Ovwah, E. (2015b). Pattern of parasitic infections in anurans from a mangrove community of the Niger Delta, Nigeria. *The Zoologist* 13: 51-56.
- Akani, G.C., Politano, E and Luiselli, L. (2004). Amphibians recorded in forest swamp areas of the Niger Delta (southeastern Nigeria), and the effects of habitat alteration from oil industry development on species richness and diversity. *Applied Herpetology* 2: 1-22.
- Akani, G.C., Luiselli, L., Amuzie, C.C. and Wokem, G.N. (2011). Helminth community structure and diet of three Afro tropical anuran species: a test of the interactive-versus-isolationist parasite communities hypothesis. *Web Ecology* 11: 11-19.
- Anyacho, O.A. (1997). Survey of helminth parasites of a common African toad *Bufo regularis* around University of Port Harcourt, Choba, Rivers State. B.Sc. thesis, Department of Zoology, University of Port Harcourt. 32pp.
- Blackburn, D. C. and Wake, D. B. (2011). Class Amphibia Gray, 1825. In: Zhang, Z.-Q. (Ed.) Animal biodiversity: An outline of higher-level classification and survey of taxonomic richness. *Zootaxa* 3148: 39-55.
- Crump, M. and Scott Jr, N. (1994). Visual Encounters Surveys. In: Measuring and Monitoring Biological Diversity-standard methods for amphibians, W. Heyer, M. Donnelly, R. McDiarmid, L. Hayek

- and M. Foster (Ed.), Smithsonian Institution Press, ISBN-10 1560982845, Washington, USA.
- Edo-Taiwo O., Ovwah E., Imasuen, A.A. and Aisien, M.S.O. (2014). Larval strigeoid trematodes in anurans from southern Nigeria. *Helminthologia* 51 (4): 318-322.
- Fowler, J. and Cohen, L. (1994). *Practical statistics for field biology*. 4<sup>th</sup> edition. John Wiley and Sons, Chichester, England.
- Imasuen, A.A. and Aisien, M.S.O. (2012). Digenetic trematodes parasitic in anurans from rainforest biotopes in Edo State, Nigeria. *The Zoologist* 10: 25-33.
- Imasuen, A.A., Aisien, M.S.O. (2015). Helminth parasites of *Silurana tropicalis* from the Okomu National Park, Edo State, Nigeria. *Nigerian Journal of Parasitology* 36(1): 61-66.
- Imasuen, A.A., Enabulele, E.E. and Aisien, M.S.O. (2012). Helminth community of tree frogs at the Okomu National Park, Edo State, Nigeria. *Nigerian Journal of Parasitology* 33: 1-8.
- Khalil, L.F., Jones, A. and Bray, R.A. (1994). *Keys to the cestode parasites of vertebrates*. International Institute of Parasitology, St. Albans, UK, 751pp.
- Kuzmin, Y. (2001): *Rhabdias africanus* sp. nov. (Nematoda, Rhabdiasidae) a new nematode species from South African toads (Amphibia, Bufonidae). *Acta Parasitologica* 46 (2): 148-150.
- Nworah D.C. and Olorunfemi, O.J. (2011). The helminth parasitofauna of *Bufo regularis* (Reuss) in Awka, Anambra State, Nigeria. *International Journal of Parasitology Research* 3: 26-30.
- Oddo, U.O. (2008). Helminth parasites of amphibians from the freshwater swamp at Sapele, Delta State, Nigeria. B.Sc. Dissertation, University of Benin, Benin City, Nigeria, pp. 53.
- Oladimeji, A.A., Abuh, S.J. and Sadiku, S.O.E. (1990). Parasitic helminths of the frog *Dicroglossus occipitalis*. *Nigerian Journal of Parasitology*, 9-11: 145-148.
- Ogoanah, S.O. (2010). Anuran species of Delta State, Nigeria, Ph.D. Thesis, University of Benin, Nigeria.
- Ogoanah, O.S., Luke, D. and Aisien, M.S.O. (2014). Anuran diversity in Mosogar, a riparian community in Delta State, Nigeria. *Nigerian Journal of Life Sciences* 4(1): 15-19.
- Omereji, A.B. (2014). *Helminth parasites of anurans from sub-urban locations in Rivers State*. B.Sc. Thesis, Rivers State University of Science and Technology, Port Harcourt. 51pp.
- Ovwah, E. (2016). *Amphibian biodiversity and helminth parasitic infections of anurans at Okomu Oil Palm Plantations, Okomu-Udo, Edo State, Nigeria*. Ph.D Thesis, University of Benin, Benin City, Nigeria
- Ozemoka, H.J. (2012). *Helminth parasites of anurans from the derived savannah at Agbede, Edo State, Nigeria*. M.Sc. Thesis, University of Benin, Benin City, Nigeria.
- Prudhoe, S. and Bray, R.A. (1982). *Platyhelminth parasites of the amphibian*. British museum (Natural History). Oxford University Press, London.
- Roedel, M. O. (2000). *Herpetofauna of West Africa*. Vol. 1: Amphibians of West African Savannah. Chimaera, Frankfurt.
- Sampson, A.S. (2014). *Helminth parasites of anurans in Rivers State University of Science and Technology*. B.Sc. Thesis, Rivers State University of Science and Technology, Port Harcourt. 35pp.

Yamaguti, S. 1971. *Synopsis of digenic trematodes of vertebrates*. Vol.1. Kaigaku Publishing Company, Tokyo.

Zar, J.H. (1984). *Biostatistical analysis*, 2<sup>nd</sup> edition. Prentice-Hall Inc. New Jersey.